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Supplementary File

Figure 4. Distribution of the obtained P. aeruginosa isolates among ICUs and different hospital departments.



Figure 5. Distribution of the obtained *P. aeruginosa* isolates among different clinical specimens



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Figure 6. Antimicrobial resistance phenotypes of the obtained *P. aeruginosa* isolates.

Figure 7. Modified Congo Red agar plate (MCRA) for detection of biofilm production among *P. aeruginosa* isolates Letter A: Dry black crystalline colonies i.e. positive for biofilm production. Letter B: Red colored colonies i.e. negative for biofilm production.



		Disk P.aeruginosa isolates (n=8				es (n=80)				
Antimicrobial groups	Antimicrobial	Abbrow	DISK		Int	erpreti	ve cate	gories		
Antimicrobiargroups	agents	ADDIEV.	(ug)		S		I		R	
			(µg)	No	%	No	%	No	%	
Penicillins	Piperacillin	PRP	100µg	29	36.3	4	5	47	58.7	
	Piperacillin-	TZP	100/10µg	38	47.5	8	10	34	42.5	
	tazobactam									
B-lactam/β-lactamase	Ceftazidime-	CZA	30/20 µg	65	81.3	0	0	15	18.7	
inhibitor combinations	avibactam									
	Ceftolozane-	C/T	30/10 µg	51	63.7	0	0	29	36.3	
	tazobactam									
Cephems	Ceftazidime	CAZ	30µg	21	26.3	0	0	59	73.8	
	Cefepime	FEP	30µg	27	33.7	0	0	53	66.3	
Monobactams	Aztreonam	ATM	30µg	25	31.3	4	5	51	63.7	
	Doripenem	DOR	10µg	25	31.3	0	0	55	68.7	
Carbapenems	Imipenem	IPM	10µg	23	28.7	1	1.3	56	70	
	Meropenem	MEM	10µg	26	32.5	0	0	54	67.5	
	Gentamicin	CN	10µg	20	25	0	0	60	75	
Aminoglycosides	Tobramycin	ТОВ	10µg	21	26.3	0	0	59	73.8	
	Amikacin	AK	30µg	30	37.5	4	5	46	57.5	
	Ciprofloxacin	CIP	5µg	40	50	0	0	40	50	
Eluoroquinolonog	Levofloxacin	LEV	5µg	37	46.3	1	1.3	42	52.4	
Fluoroquinoiones	Norfloxacin	NOR	10µg	47	58.7	0	0	33	41.3	
	Ofloxacin	OFX	5μg	45	56.3	0	0	35	43.7	
Phosphonic acid	Fosfomycin	FF	50µg	50	62.5	0	0	30	37.5	

Table 4. Antimicrobial susceptibility pattern of the obtained *P.aeruginosa* isolates (n=80) by disk diffusion method (CLSI, 2022).

		P. ae	<i>ruginosa</i> isolates (r	1=80)	<i>Kappa</i> <i>agreement</i> p-value			
Phenotypic tests		Combined di test (CDT	sk confirmatory Γ) for ESβLs	Total				
		+ve (n=10)	-ve (n= 70)					
Disk	+ve (n=59)	9 (15.3%)	50 (84.7%)	59 (100%)	K1 = 0.060*	p1 <0.001*		
diffusion	-ve(n=21)	1 (4.8%)	20 (95.2%)	21 (100%)				
screening								
test								
•		AmpC disk o	confirmatory test					
		+ve (n=37)	-ve (n=43)					
				Total				
	+ve (n=74)			74 (100%)				
Cefoxitin								
disk		35 (47.3%)	39 (52.7%)					
diffusion		, , , , , , , , , , , , , , , , , , , ,	· · · · · · · · · · · · · · · · · · ·		<i>K</i> 2= 0.036*	p2 <0.001*		
screening	-ve (n=6)	2 (33.3%)	4 (67.7%)	6 (100%)		*		
method								

Table 5. Agreement between phenotypic screening and confirmatory methods for $ES\beta Ls$ and AmpC production among *P.aeruginosa* isolates

**K*1: Kappa forESβLs*K*2: Kappa for AmpC*: Statistically significant

Kappa interpretation:

Kappa < 0: No agreement

Kappa between 0.00 and 0.20: Slight agreement

Kappa between 0.21 and 0.40: Fair agreement

Kappa between 0.41 and 0.60: Moderate agreement

Kappa between 0.61 and 0.80: Substantial agreement

Kappa between 0.81 and 1.00: Almost perfect agreement.

			P. aerug	<i>inosa</i> isolates	(n=80)		Kappa agreement	p-value
Phenotypic tests		Imipenen acid CD te A carbap (KI	n/boronic st for class enemases ?C)	Imipenem/EDTA CD test for class B carbapenemases (MBL)Total			<i>K</i> 1= 0.141	p1= 0.013*
		+ve (n=12)	-ve (n=68)	+ve (n=31)	-ve (n=49)			
Disk diffusion	+ve (n=56)	12 (21.4%)	44 (78.6%)	31(55.4%)	25(44.6%)	56 (100%)		
screening test	-ve (n= 24)	0 (0%)	24 (100%)	0 (0%)	24 (100%)	24 (100%)	<i>K</i> 2= 0.427	p2<0.001*

CD: combined disk

Table 6. Frequency of class A and class B carbapenemases production among *P. aeruginosa* isolates by phenotypic screening and confirmatory methods

*K*1: Kappa for KPC

p1: p-value for KPC p2: p-value for MBL

*: Statistically significant

Kappa interpretation:

Kappa < 0: No agreement

Kappa between 0.00 and 0.20: Slight agreement

Kappa between 0.21 and 0.40: Fair agreement

Kappa between 0.41 and 0.60: Moderate agreement

Kappa between 0.61 and 0.80: Substantial agreement

Kappa between 0.81 and 1.00: Almost perfect agreement.

*K*2: Kappa for MBL

Table 7. Susceptibility pattern of MDR and XDR *P. aeruginosa* isolates to the new beta-lactam/beta-lactamase inhibitor combinations: ceftazidime-avibactam and ceftolozane-tazobactam by disk diffusion method

P. aeruginosa	Ceftazidime-avibactam (Disk diffusion)			actam n)	Z	n-value	Ceft	olozane• (Disk di	-tazoba ffusion)	ctam)	Z	p-value
		S		R		P	S		S R			P
	No.	%	No.	%			No.	%	No.	%	-	
MDR isolates (n=26)	21	80.8	5	19.2	4.16	<0.001*	17	65.4	9	34.6	1.94	0.052
XDR isolates (n=42)	32	76.2	10	23.8	3.60	<0.001*	22	52.4	20	47.6	0.22	0.825

*: Statistically significant

Table 8. Correlation between antimicrobial resistance phenotypes and biofilm formation among *P. aeruginosa* isolates

Biofilm formation	N P. aer (n	MDR P. aeruginosa (n=26)		XDR P. aeruginosa (n=42)		Non-MDR P. aeruginosa (n=12)		Total (n=80)		p-value
	No.	%	No.	%	No.	%	No.	%		
Yes (n=47)	13	50	34	81	0	0	47	58.8	28.205	<0.001*
No (n= 33)	13	50	8	19	12	100	33	41.2		

Characteristics	Colistin- isola	resistant ates	Colistin– non resistant isolates		χ2	p-value
	(n=15) (n=65)					
	No.	%	No.	%		
Hospitalization in ICU	10	66.7	23	35.4	Z= 4.23	<0.001*
**Prolonged hospitalization>21 days	9	60	6	9.2	Z= 4.18	<0.001*
Invasive procedures	15	100	65	100	21.48	<0.001*
Co-morbidities						
Diabetes mellitusHypertension	12	80	45	69.2		
COPD****Chronic renal failure	0	0	2	3.1		
Chronic liver diseasesMalignancy	0	0	14	21.5		
Combined comorbidities	5	33.3	11	16.9	0.69	0.406
	0	0	0	0		
	0	0	0	0		
	4	26.7	7	10.8		
	3	20	11	16.9		
VAP***	7	46.7	6	9.2	Z=3.550	0.004*
Prior antibiotic exposure	15	100	65	100	Z=1.147	0.250
Prior colistin therapy						
Monotherapy Combination therapy	15	100	5	7.7	16.04	~0.001*
• Combination incrapy	0	0	12	18.5	10.94	<0.001*

Table 9. Univariate analysis of different risk factors associated with colistin-resistant P. aeruginosa isolates infection