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Seroprevalence and risk factors of foot-and-mouth disease in sheep and goats in the Soudan-Sahelian and Guinean regions of Cameroon

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ABSTRACT

Background: Information on the epidemiology of foot and mouth disease (FMD) in small ruminants in Cameroon and particularly in the northern regions is insufficient. Indeed, studies on the epidemiology of FMD in these areas were focused on cattle and pigs. Hence, the current cross-sectional study was conducted from January to July 2020 to determine the seroprevalence and risk factors of FMD in small ruminants and cattle mixed herds in the Soudan-Sahelian and Guinean agro-ecological zones (AEZs) of Cameroon. **Methods**: A total of 350 serum samples from 268 sheep and 82 goats were collected in the 3 regions (Adamawa (n=93), North (n=105), and Far North (n=152)). All samples were tested using indirect ELISA specific to antibodies against FMD virus Non-structural Protein (NSP) and seropositive samples were further serotyped using ELISA "PrioCHECK® FMDV Type A" and "ELISA PrioCHECK® FMDV Type O" kits. Moreover, a univariate analysis was performed to test the association between the potential risk factors and prevalence of FMD. Lastly, a multivariate logistic regression was used to build the final model. Results: The overall seroprevalence of antibodies to the non-structural protein of the FMD virus was 45.4% (95% CI: 40.3 - 50.7), that of serotype A was 4.6% (95% CI: 2.8 - 7.3) and that of serotype O was 17.4% (95% CI: 13.8 - 18.7). NSP antibodies seroprevalence was significantly higher in sheep, but anti-serotype O antibodies were higher in goats. The main risk factor independent of the species is region. In fact, small ruminants from the Sahelian region (OR: 7.3; P=0.003) had a higher seroprevalence than others. No intrinsic factor showed a significant influence on seroprevalence in goats. However, in sheep, age, sex and physiological status were significantly associated to seroprevalence. For both species, young animals (OR: 2.9; P=0.02) were more susceptible, females were 1.4 and 4.7 times respectively more likely to have NSP and type O antibodies than males. This study has shown that FMDV antibodies are present and the viruses could be circulating among small ruminant populations in the three regions of Cameroon. Conclusion: The present study highlights the probable circulation of FMD serotypes A and O in both sheep and goat populations for the first time in the major cattle rearing northern regions of Cameroon. In depth molecular studies on the serotypes circulating in these regions are therefore needed to identify potential vaccine candidates and to design species-specific vaccines. Ultimately, the real burden and economic impact of this disease should be assessed nationwide.

Introduction

Foot-and-mouth disease (FMD) is a highly contagious viral disease of cloven-hoofed ungulates (Artiodactyls) of both domestic and wild animals affecting mostly bovine, ovine, caprine and porcine species [1]. Foot-and-mouth disease is caused by a positive-sense, single stranded RNA virus of the **Aphthovirus** within the family genus Picornaviridae. Seven serotypes of FMDV have already been reported (O, A, C, Asia-1, SAT1-3), with numerous antigenically distinct subtypes within each serotype [2,3]. These serotypes have further been classified into topotypes, which are based on geographic region as well as on the VP1 sequence-based phylogenetic analysis. There is no lasting cross-protection between serotypes and animals having an immune response against one serotype (by either vaccination or infection) might not be protected against the other serotypes [4]. Furthermore, vaccination programs require vaccines that match the circulating viral strains [5]. The clinical signs of FMD include fever, lameness, and salivation (ptyalism) associated with the appearance of characteristic vesicular lesions in the oral cavity as well as interdigital clefts and coronary bands of the feet. Though mortality rates are generally low among adult animals, production objectives such as weight gain, milk yield, and draught power are severely affected by the clinical disease [6].

Foot-and-mouth disease constitutes a menace to the livestock sector in many countries of the world due to its high contagiousness and the threat it poses to international trade of livestock and livestock products [5,7]. It is one of the most important economic diseases of cattle in the tropics limiting animal production and trade and contributes towards food insecurity in these regions [8]. Lack of infrastructure, economic resources and vaccines tailored to particular condition is rendering many developing countries vulnerable to the spread and devastating effects of the disease. In Africa, >75% of livestock are reared under communal smallholder systems [9] which are most negatively impacted by animal diseases including transboundary animal diseases (TADs) such as FMD.

The disease is enzootic in Cameroon and most parts of Africa. In Cameroon, high rates of serotypes O and A have been reported in the North West, Adamawa, North and Far North regions [10-13] and serotype SAT2 has been frequently detected in the Adamawa and Far North regions [11-13]. Serotypes SAT1 and SAT3 have also been reported at low levels in the country [13]. There is no formal FMD control program in Cameroon because international trade of livestock is not a priority. However, in March 2014, cattle vaccination against FMD was launched in the country using a commercially available trivalent inactivated FMDV vaccine against serotypes SAT2, A, and O [14]. Although the virus strains used in the vaccine were based on vaccine matching tests with field isolates, it is important to note that the vaccine prevented clinical manifestation and not the persistence of the infection [15].

Despite several FMD research in livestock [10-13, 16-18], the virus ecology and maintenance during inter-epidemics is not well understood [13] and there is dearth of information of the disease in small ruminants in Cameroon. Chepkwony et al. [19] reported that small ruminants function as silent repository for FMD virus, and are responsible for inter-epidemic survival of FMD virus and spill over to large ruminants or other susceptible animal species and could cause repeated FMD outbreaks.

Though small ruminants and cattle usually occupy the same microenvironments and are grazed together [13], in Cameroon, the epidemiology of FMD is focused on cattle and pigs [20-22]. There is no study on the epidemiology of FMD in small ruminants in the North region of Cameroon. Given that small ruminants do not frequently present symptoms, which make the clinical detection of the disease difficult [23], it is important to investigate on the role of small ruminants in FMD epidemiology. Non-structural proteins are formed during replication of the virus and antibodies to these proteins are only present in animals infected with the live virus [24]. In this context, the present conducted determine study to seroprevalence and associated risks factors of FMD

in domestic small ruminants reared with large ruminants in the Northern regions of Cameroon.

Materials and methods

Study area

A cross-sectional study was carried out in Adamawa $(7^{\circ}09' - 7^{\circ}70'N \text{ and } 13^{\circ}52' - 13^{\circ}70'E)$, North $(8^{\circ} - 10^{\circ} \text{ N and } 12^{\circ}27' - 15^{\circ}62' \text{ E})$ and Far North (10° - 12° N and 14° - 15° E) regions of Cameroon (Figure 1). The Adamawa region is a Savannah-Guinean highland while the North and Far North regions are typical Sudan-Sahelian agroecological zones located in the mid to high altitude zones of Cameroon. The Adamawa region has an annual precipitation of 1200 - 1600 mm, rainy season from about mid-March to October and temperatures of 14° C- 26°C while the North and Far North regions has annual precipitations of 400 – 900mm, rainy season from July to October and temperatures of 21°C - 36°C [25]. These three northern regions represent the major centers of small ruminants farming in Cameroon [26]. The Adamawa region is a Carrefour between the northern and southern parts of Cameroon and provides the only road access to the North and Far North regions from the southern regions and viceversa for many socioeconomic ventures [26]. The major livestock rearing regions of Cameroon are the Far North, Adamawa, North and North West. The livestock population of Cameroon includes 31 million poultry, 6 million cattle, 7 million small ruminants, one million pigs, 150,000 donkeys and 15,000 horses. The proportion of cattle by administrative region of Cameroon is as follows: 37.5% in the Far North, 33.9% in Adamawa, 11.6% in the North, 8% in the North West, 6.3% in the East and 2.7% in the West [14]. Most small ruminants are produced in the three regions of the North, Far North and Adamawa (>70% for sheep and approximately 60% for goats). The livestock population in the Northern regions of Cameroon is a mixture of sedentary (village) and mobile (seasonally transhumant) animals [26]. In general, the sedentary herds are smaller but more numerous and may contain sheep, goats, cattle and pigs, while the mobile herds are larger but fewer in number and contain cattle, sheep and goats. These two production systems share pasture and water resources as well as trade networks (markets) and veterinary services. In addition, livestock is the main source of income for about 30% of the rural population in northern Cameroon. The insecurity situation in neighbouring countries (Nigeria, Chad and CAR) has led to the massive displacement of livestock farmers and their livestock mainly to the North and Adamawa regions, constituting to a high risk for the introduction and spread of diseases. Thus, the northern region constitutes a hub for livestock breeding in the Central African Subregion.

Sample size determination

The small and large ruminant samples were collected from the three study regions (North, Far North and Adamawa) and this sample number depended on the small ruminant population in these regions. 43.4% small ruminants were sampled in the Far North region, 30% in the North region and 26.6% in Adamawa region. The selection of herds was conducted by using the random-number generation method of known ruminant owners and locations of herds listed at the regional delegations of the Ministry of Livestock, Fisheries and Animal Industries (MINEPIA). Farms where cattle and small ruminants were reared together in areas where FMD has previously been reported were sampled. A total of 51 farms fitting the criteria and willing to participate in the study were surveyed with the help of MINEPIA field staff. Based on the 27.84% prevalence obtained by Lazarus et al. [27], a minimum number of 373 small ruminants to be sampled was calculated according to Thrusfield [28] with a significance threshold of 95% and required precision of 5%. Selection of small ruminants within chosen herds was based on a systematic random sampling technique and the willingness of the owner. The sample size was rounded up to 350 samples for the 51 farms to avoid sampling bias.

Blood collection, processing and storage

Blood was collected in 5ml vacutainer tubes via the jugular vein puncture of small ruminants used in the study. Information on the animals selected (age, sex, body condition score, breed and physiological status) were registered on the identification sheets and each sample was identified with a unique code. Serum was extracted, identified with the same sample code from tube with whole blood and transported in ice-boxes, fitted with icepacks, and transported to the National Veterinary Laboratory (LANAVET), Garoua-Cameroon and stored at -20°C prior to ELISA testing.

Ethics and approval

Prior to sampling, the research approval for this study was obtained from the Ministry of Livestock, Fisheries and Animal Industries (MINEPIA) and the National Veterinary Laboratory (LANAVET). Other administrative approvals required for the study were obtained from the Services of Livestock, Fisheries and Animal Industries at the regional and district levels. Each owner of a herd selected for sampling provided verbal consent, once the objectives of the study were clearly explained in a language they best understood and all their questions and doubts addressed accordingly. Herds whose owners did not consent were replaced with the next herd in the random sample list.

Characteristics of selected animals

The age was provided by the owner and/or determined by dental observation [29] while the Body Condition Score (BCS) on a scale of 0 to 5 was determined as previously described by Hervieu et al. Q1[29]. Small ruminants were grouped into three (3) age classes as proposed by Aklobessi [31] as such: young (age ≤ 1 year); adults (animals in production stage) age > 1 year and ≤ 3 years) and old animals or animals at the end of their production stage (age> 3 years). Similarly, a classification according to the size of the herd was carried out as follows: small herd (size ≤ 10 animals); medium herd (10 to 20 animals) and large herd (≥ 20 animals).

Detection of antibodies against FMD virus nonstructural proteins

An indirect ELISA was used for the detection of antibodies directed against the non-structural proteins of FMD virus, irrespective of the serotype involved and the vaccination status of the animal [32]. In this study, "PrioCHECK® FMDV-NS" Kit (Pronics Lelystad B.V. the Netherlands) was used according to the protocol provided by the manufacturer.

Detection of antibodies against serotypes A and O

The Non-structural protein ELISA positive samples were retested for A and O serotyping. The ELISA "PrioCHECK® FMDV Type A" and "ELISA PrioCHECK® FMDV Type O" kits were used to detect antibodies against FMDV A and O serotypes [33] according to the protocol provided by the manufacturer.

Statistical analysis

Individual animal laboratory data generated during testing alongside individual animal biodata obtained during sample collection (species, sex, age) were entered in Microsoft Excel

(Microsoft Corporation. (2018). Data in MS Excel was cleaned and coded before being exported and analysed using IBM Statistical Package for Social Sciences (SPSS) software IBM Corp. Released 2015. IBM SPSS Statistics for Windows, Version 23.0. Armonk, NY: IBM Corp. Descriptive analysis was used to determine the seroprevalence of FMD based on endogenous and exogenous factors. A univariate analysis using the χ 2 test and Fischer test (when the conditions of the χ 2 test were not met) was performed to test the association between the potential risk factors and seroprevalence of FMD as recommended by Campbell [34] and Richardson [35]. Factors with p<0.2 after univariate analysis were introduced into the final model. A multivariate logistic regression was then used to build the final model with seropositivity to NSP; type A; type O as the dependent variable and risk factors as independent variables. The odds ratios of the risk factors with their respective 95% confidence intervals (CI) were obtained in the final model. A value of P <0.05 was considered as significant association in this study.

Results

The characteristics of the sampled animal species are presented in **Table 1**.

Anti-FMD seroprevalence

The anti-FMDV antibodies are frequent in small ruminants (anti-FMDV-NSP antibodies (45.4% (95% CI: 40.3 – 50.7)). Furthermore, anti-FMDV Serotype A (4.6% (95% CI: 2.8 – 7.3)) and anti-FMDV Serotype O (17.4% (95% CI: 13.8 - 21.8)) were widespread in the Northern regions of Cameroon (**Table 2**). Overall, 45.4% (159) of 350 small ruminants tested in the three regions (Adamawa (n=93), North (n=105), Far North (n=152)) showed positive reactions for at least one of the anti-FMDV antibodies detection method used in this study.

Risk factors of FMD in the study area

In both species (**Table 3**), region was significantly associated to FMDV seroprevalence. In sheep, old animals and pregnant females recorded highest seroprevalences for NSP anti-antibodies while the physiological status was not a risk factor for type O. In Goats, none of the tested parameters was significantly linked to any of the estimated antibody seroprevalence.

When the two species were considered together, the main risk factor was locality (**Table 4**). In fact, animals in the Far North region were more

likely to be exposed to type A (OR: 7.3 (95% CI: 1.9 - 27.9), p=0,004) than in the North region. Young animals are more likely to have FMD as per the NSP (OR: 2.2 (95% CI: 1.0 - 4.7), P=0.049) and type O

(OR=4.8 (95% CI: 1.7 – 13.7), P=0.003) seroprevalences. There was no significant difference for the other parameters.

Table 1. Characteristics of small goats and sheep populations sampled in the three northern regions of Cameroon in 2020.

Parameters		Species		Total	
		Goats	Sheep		
Sex	Female	64	191	255	
	Male	18	77	95	
Age	Adult	51	118	169	
	Young	17	90	107	
	Old	14	60	74	
Body condition	Fat	1	2	3	
score	Thin	22	78	100	
	Normal	59	188	247	
Breed	Djalonke	33	50	83	
	Kirdi	24	77	101	
	Mayo kebi	0	42	42	
	Oudah	0	57	57	
	Peuhl	0	14	14	
	Rousse	1	0	1	
	Sahel	24	1	25	
	Woïla	0	27	27	
Total	•	82	268	350	

Table 2. FMD NSP antibodies, serotypes A and O prevalence in small ruminants (sheep and goats) of the northern regions of Cameroon in 2020.

Parameters	Detection of anti-FMDV antibodies								
	Anti-NSP anti	bodies	Anti-Type A	antibodies	Anti-Type O antibodies				
	N (%)	p (χ2)	N (%)	p (χ2)	N (%)	p (χ2)			
Region		<0.0001 (75.6)		< 0.0001		< 0.0001			
Adamawa (93)	78 (83.9)		0 (0)	(21.5)	0 (0)	(26.8)			
Far North (152)	49 (32.2)		3 (2.0)		35 (23.0)				
North (105)	32 (30.5)		13 (12.4)		26 (17.1)				
Species		<0.0001 (13.9)		0.233		0.004 (8.3)			
Goats (82)	52 (63.4)		2 (2.4)	(1.16)	23 (28.0)				
Sheep (268)	107 (39.9)		14 (5.2)		38 (14.2)				
Age group		0.003 (11.6)		0.475 (1.5)		0.002 (12.6)			
Young (107)	34 (31.8)		3 (2.8)		8 (7.5)				
Adults (169)	87 (51.5)		10 (5.9)		33 (19.5)				
Old(74)	38 (51.4)		3 (4.1)		20 (20.7)				
Body condition score		0.275 (2.8)		0.812(0.2)		0.129 (4.7)			
Thin (100)	48 (48.0)		5 (5.0)		24 (24.0)				
Normal (247)	111 (44.9)		11 (4.5)		37 (15.0)				
Fat(3)	0		0		0				
Sex		0.002 (8.6)		0.326 (0.6)		< 0.0001			
Female (255)	128 (50.2)		13 (5.1)		50 (22.0)	(13.4)			
Male (95)	31 (32.6)		3 (3.2)		6 (5.3)				
Total (350)	159 (45.4)		16 (4.6)		61 (17.4)				
(95% IC)	(40.3 - 50.7)		(2.8 - 7.3)		(13.8 -				
					21.8)				

FMDV = Foot and mouth disease virus, NSP = Non-structural protein of FMDV, Type A = FMDV serotype A, Type O = FMDV serotype O

Table 3. Seroprevalence and associated risk factors of FMDV in small ruminants of the northern regions of Cameroon in 2020.

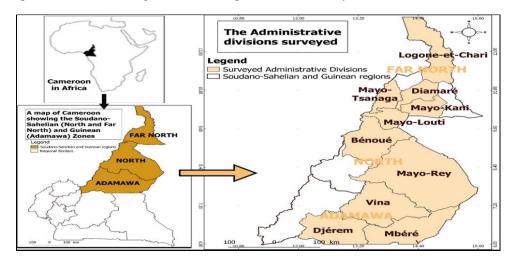
III 2020.	Detection of anti-FMDV antibodies in Sheep#(n =268)					Detection of anti-FMDV antibodies in Goats (n=82)						
Parameters		-NSP		Type A	Anti-T		Anti-		Anti-T		Anti-	Type O
		odies		odies	antib	odies	antib	odies	antibo	odies		bodies
	N (%)	p (χ2)	N (%)	p (χ2)	N (%)	p (χ2)	N (%)	p (χ2)	N (%)	p (χ2)	N (%)	p (χ2)
Region		< 0.0001		< 0.0001		0.001		0.018		0.51		< 0.0001
Adamawa	51	(65.8)	0 (0.0)	(18.3)	0	(13.0)	27	(8.1)	0	(1.4)	0	(22.2)
Far North	(85.0)		2 (1.6)		(0.0)		(81.8)		(0.0)		(0.0)	
North	36		12		22		13		1		13	
	(28.3)		(14.8)		(17.3)		(52.0)		(4.0)		(52.0)	
	20				16		12		1 (4.2)		10	
A = = = =====	(24.7)	0.01		0.23	(19.8)	0.001	(50.0)	0.61	(4.2)	1	(41.7)	0.94
Age group Young	25	(8.4)	2 (2.2)	(2.9)	4	(14.8)	9	(1.2)	1	(1.3)	4	(0.2)
Adults	(27.8)	(0.4)	9 (7.6)	(2.9)	(4.4)	(14.6)	(52.9)	(1.2)	(5.9)	(1.3)	(23.5)	(0.2)
Old	54		3 (2.0)		18		33		1		15	
Old	(45.8)		3 (2.0)		(15.3)		(64.7)		(2.0)		(29.4)	
	28				16		10		0		4	
	(46.7)				(26.7)		(71.4)		(0.0)		(28.6)	
Body		0.30		1.00	, ,	0.14	, ,	0.19	` /	1	, ,	0.57
condition	36	(2.6)	4 (5.1)	(0.8)	16	(3.6)	12	(3.0)	1	(2.9)	8	(1.4)
score	(46.2)		10		(20.5)		(54.5)		(4.5)		(25.4)	
Thin	71		(5.3)		22		40		1		15	
Normal	(37.8)		0		(11.7)		(67.8)		(1.7)		(36.4)	
Fat	0 (0.0)				0		0		0		0	
Sex	0.4	0.02		0.39	2.5	0.004	4.4	0.05		0.61	2.1	
Female	84	(4.6)	11	(0.4)	35	0.001	44	(3.6)	2	(1.0)	21	
Male	(44.0)		(5.8)		(18.3)	(9.4)	(68.8)		(3.1)		(32.8)	
	23 (29.9)		3 (3.9)		(3.9)		8 (44.4)		0		2 (11.1)	
Physiological	(29.9)				(3.9)		(44.4)				(11.1)	0.98
status	24	0.005	2 (3.2)	0.28	11	0.59	20	0.36	1	0.29	9	(0.04)
Nursing	(38.1)	(10.6)	3 (4.6)	(2.6)	(17.5)	(1.04)	(74.1)	(2.0)	(3.7)	(2.5)	(33.3)	(0.01)
Pregnant	39	(====)	6 (9.5)	(=10)	10	(=10-1)	19	(=10)	0	(=)	9	0.31
Empty	(60.0)		, ,		(15.4)		(70.3)		1		(33.3)	(1.0)
1.7	21	-	-		14	-	5	0.38	(10.0)	-	3	, ,
Castrated	(33.3)		-	-	(22.2)		(50.0)	(0.8)			(30.0)	
Whole									0			
	-				-		1		0		1	
	-				-		(25.0)				(25.0)	
							(50.0)				(7.1)	
Total	107		14		38		52		2		23	
(95% IC)	(39.9)		(5.2)		(14.2)		(63.4)		(2.4)		(28.0)	
	(33.9 –		(3.1 –		(10.4		(15.0		(0.2 –		(5.7 –	
	45.6)		8.5)		_		_		2.7)		12.6)	
					18.7)		24.4)					

FMDV = Foot and mouth disease virus, NSP = Non-structural protein of FMDV, Type A = FMDV serotype A, Type O = FMDV serotype O. # Physiological status computed only for females.

Table 4. Multivariable logistic regression model of FMDV associated risk factors in small ruminants of the northern regions of Cameroon, 2020.

Parameters		Detection of anti-FMDV antibodies								
		NSP			Type A	Type O				
		P-value OR (95% CI)		P-value	OR (95% CI)	P-value	OR (95% CI)			
Region	Adamawa	0.0001	0.06(0.03-0.15)	0.994	/	0.992	/			
	Far North	0.781	0.9 (0.5 – 1.7	0.004	7.3 (1.9 – 27.9)	0.703	1.1 (0.6 - 2.2)			
	North		Ref	•	Ref		Ref			
Species	Goat	0.022	0.5(0.3-0.9)	0.393	1.9 (0.4 – 9.7)	0.0001	0.2 (0.1 - 0.5)			
	Sheep		Ref	•	Ref		Ref			
Sex	Female	0.016	0.3 (0.1 – 0.8)	0.452	0.6 (0.1 – 2.5)	0.021	0.2 (0.07 – 0.8)			
	Male		Ref	•	Ref		Ref			
Age	Adult	0.368	1.3 (0.7- 2.5)	0.685	0.7(0.2-3.0)	0.257	1.5 (0.7 – 3.2)			
group	Young	0.049	2.2 (1.0 - 4.7)	0.468	1.9 (0.3 – 11.1)	0.003	4.8 (1.7 – 13.7)			
	Old	•	Ref	•	Ref	•	Ref			
Body	Fat		/		/		/			
Condition	Thin	0.420	0.8 (0.5 – 1.4)	0.297	0.5 (0.2 – 1.7)	0.027	0.5(0.2-0.9)			
Score	Normal	•		ė	Ref		Ref			
Physiolog	Nursing	0.779	1.1(0.5-2.4)	0.218	2.5 (0.6 – 11.2)	0.196	1.8 (0.7 – 4.3)			
ical Status	Pregnant	0.557	2.4 (0.1 – 42.1)	0.332	2.1 (0.5 – 9.1)	0.591	1.3 (0.5 – 3.1)			
	Empty		Ref	•	Ref		Ref			

Figure 1. Map of Cameroon showing the Northern regions with the surveyed administrative divisions.



Discussion

The current study aimed at determining the seroprevalence and risk factors of FMD in small ruminants and cattle mixed herds in the Soudan-Sahelian and Guinean agro-ecological zones (AEZs) of Cameroon. We found that the percentage of positive sera for each serotype was different in sheep and goats in the current study. Non-Structural Protein (NSP) antibodies were found in 159 animals (45.4%), indicating that the animals had come into

contact with the virus (presumably the wild strain) since they were not vaccinated. Serotype A antibodies were the least common in both sheep and goats. These findings are different from those reported by Habiela et al. [38], where serotype A was the most prevalent followed by serotype O. The later was the most prevalent in another survey conducted in small ruminants [37]. The distribution of these serotypes is different for both regions (North and Far North) of the Soudan Sahelian agroecological zone. None of the serotypes targeted were

detected in the Guinean region (Adamawa), probably because of the breeding system, where less breeders raise small ruminants with bovines.

The free movements of animals and coexistence with other ruminants in the same habitat, mainly cattle, may contribute to cross transmission of FMDV. In addition, it is advisable to control FMD outbreaks by restricting movement of small ruminants for two weeks after infection as sheep and goats are involved in the transmission of FMDV during the early stages of the disease [38]. Therefore, small ruminants should be included in the vaccination program [36]. It is worth noting that no animal related variables showed significant relationship with seropositivity. Husbandry related variables showed significant relationship with seropositivity in studies of Balinda et al. [39] and Chepkwony et al. [19] in Kenya.

At individual animal level, a significant difference was observed in seroprevalence of FMD among adults and young sheep and goats. This is in agreement with the results of Habiela et al. [38] even though the seropositivity levels in our study were higher. The difference in seropositivity between age groups may be due to the fact that mature animals might have experienced several exposures to FMD at grazing fields, watering points and at market places than in age group less than one year. Therefore, adult animals might have acquired infection from multiple strains and serotypes thus producing antibodies against multiple virus incursions of FMD. The low seroprevalence in young animals might also be indicative of persistent passive immunity and less frequency of exposure of the animal to the disease as the farmers keep their lambs and kids at homesteads.

Conclusion

This study was carried out to determine the seroprevalence and associated risk factors of FMD in small ruminants reared with cattle in the Soudan-Sahelian and Guinean regions of Cameroon. The overall seroprevalence of antibodies to the non-structural protein of the FMD virus was 45.4%, that of serotype A was 4.6% and that of serotype O was 17.4%. In sheep, age, sex and physiological status were significantly associated to seroprevalence. It is advisable that SATs (SAT1, 2& 3) serotyping be conducted using the current study samples. Molecular studies are required for more insights on the strains circulating in these regions for the elaboration of species-specific vaccines and

detection of potential vaccine candidates. Ultimately, the real economic impact of this disease should be assessed nationwide.

Authors' contributions

SDJ, JAN, RNG, ID, LGY, SLS, CMM, ATN, AW: Study design, sample collection, statistical analysis, and drafted the manuscript. All authors have read, reviewed, and approved the final manuscript.

Data availability

The data used to support this study are available upon request.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- 1- Nikiforov V, Shcherbakov A, Chvala I, Kremenchugskaya S, Korennoy F, Mayorova T, Timina A, Tyulegenov S, Abdrakhmanov S, and Berdikulov M. Insights into the Molecular Epidemiology of Foot-and-Mouth Disease Virus in Russia, Kazakhstan, and Mongolia in Terms of O/ME-SA/Ind-2001e Sublineage Expansion. Viruses 2023; 15: 598.
- 2- Ranaweera LT, Kumari Wijesundara U, Jayarathne HS, Knowles N, Wadsworth J, Mioulet V. Characterization of the FMDV serotype-O isolates collected during 1962 and 1997 discloses new topotypes, CEY-1 and WCSA-1, and six new lineages. Sci. Rep 2019; 9: 14526.

- 3- Lycett S, Tanya VN, Hall M, King DP, Mazeri S, Mioulet V, Knowles NJ, Wadsworth J. The evolution and phylodynamics of serotype A and SAT2 foot-and-mouth disease viruses in endemic regions of Africa. Sci. Rep 2019; 9: 5614.
- 4- Gubbins S, Paton DJ, Dekker, A, Ludi AB, Wilsden G, Browning CFJ, Eschbaumer M, Barnabei J, Duque H, Pauszek LL, King DP. Predicting cross-protection against foot-andmouth disease virus strains by serology after vaccination. Front. Vet. Sci 2022; 9: 1027006.
- 5- Willems T, Annebel De V, Perez-Filgueira M, Yanmin Li, Ludi A, Lefebvre D, Ginette W, Bob S, Bernd H, Nora M. FMD vaccine matching: Inter laboratory study for improved understanding of r1 values. J. Virol. Methods 2020; 276: 113786.
- 6- Buckle K, Bueno R, McFadden A, Van Andel M, Spence R, Hamill C. Detection of Footand-Mouth Disease Virus in the Absence of Clinical Disease in Cattle and Buffalo in South East Asia. Front. Vet. Sci 2021; 8: 691308.
- 7- Panel NB, Alhaji J, Amin MB, Aliyu B, Mohammad OO. Economic impact assessment of foot-and-mouth disease burden and control in pastoral local dairy cattle production systems in Northern Nigeria: A cross-sectional survey. Prevent. Vet. Med 2020; 177: 104974.
- 8- Chanchaidechachai T, Saatkamp H, Inchaisri C, Hogeveen H. Analysis of Epidemiological and Economic Impact of Foot-and-Mouth Disease Outbreaks in Four District Areas in Thailand. Front. Vet. Sci 2022; 9: 904630.
- 9- Marshal K, John P, Gibson OM, Mwacharo JM, Aynalem H, Tesfaye G. Livestock Genomics for Developing Countries African Examples in Practice. Front. Genet. Sec. Livestock Genomics 2019; 10.

- 10-Lendzele SS, Koumba AA, Rodrigue MN, Mavoungou JF. Foot and Mouth Disease in Cameroon: A Systematic Review to Support its Progressive Control. J. Clin. Vet. Res 2022; 2.
- 11-Bronsvoort BMdC, Nfon C, Hamman SM, Tanya VN, Kitching RP, Morgan KL. Risk factors for herdsman-reported foot-and-mouth disease in the Adamawa province of Cameroon. Prevent. Vet. Med 2004b; 66: 127-139.
- 12-Lendzele SS, Armel KA, Rodrigue MN, Francois MJ, Burinyuy KA. Clinical Footand-Mouth Disease in Non-Vaccinated Smallholder Dairy Cattle in Adamawa Region, Cameroon: Prevalence, Farmer's Knowledge and Practices. J. Vet. Med. Surg 2021; 5: 003.
- 13- Ehizibolo DO, Fish IH, Brito B. Characterization of transboundary foot-and-mouth disease viruses in Nigeria and Cameroon during 2016. Transbound. Emerg. Dis 2020; 00: 1–14.
- 14-Sevidzem SL, Mavoungou JF, Mintsa NR. Veterinary Pharmaceuticals Sold in Cattle Markets for the Management of Foot-and-Mouth Disease and Flies in Vina Division (Adamawa-Cameroon). Dairy Vet. Sci. J 2020; 10: 555782.
- 15-Miranda RB, Delgado A, Pauszek SJ. Effect of vaccination on cattle subclinically infected with foot-and-mouth disease virus in Cameroon. Prevent. Vet. Med 2018; 155: 1-10.
- 16-Sevidzem SL, Mamoudou A, Mavoungou JF, Ikoum D. Serological Epidemiology of Footand-mouth Disease among Sedentary Mixedspecies Herds in Adamawa Region, Cameroon. J. Adv. Microbiol 2019a; 1-14.
- 17-Sevidzem SL, Mamoudou A, Dickmu S, RenzA. Risk Factors for the Contamination of WildStomoxys niger niger Macquart 1851 (Diptera:

- Muscidae) with the Foot-and-Mouth Disease Virus", Curr. Res. Agric. Sci 2019b; 6(2): 95-108.
- 18- Lendzele SS, Mavoungou JF, Kong AB, Koumba AA. Efficacy and application of a novel topical anaesthetic wound formulation for treating cattle with Foot-and-Mouth disease: a field trial in Cameroon. Transbound. Emerg. Dis 2020; 1–12.
- 19-Chepkwony EC, Gitao GC, Muchemi GM, Sangula AK, Kairu-Wanyoike SW. Epidemiological study on foot-and-mouth disease in small ruminants: Sero-prevalence and risk factor assessment in Kenya. PLoS One 2021; 16(8): e0234286.
- 20-Bronsvoort BMD, Radford AD, Tanya VN, Nfon C. Molecular epidemiology of foot-andmouth disease viruses in the Adamawa Province of Cameroon. J. Clin. Microbiol 2004a; 42: 2186-2196.
- 21- Pomeroy LW, Bjørnstad ON, Kim H, Jumbo SD. Serotype-Specific Transmission and Waning Immunity of Endemic Foot-and-Mouth Disease Virus in Cameroon. PLoS ONE 2015; 10 (9): e0136642.
- 22-Dickmu SJ, Awah-Ndukum J, Aziwo NT Sevidzem SL. Molecular and Serological Epidemiology of Foot-and-Mouth Disease Virus in North Region of Cameroon", Adv. Microbiol 2022; 12: 579-595.
- 23-Muthukrishnan M, Singanallur BN, Villuppanoor AS. Experimental Infection of Foot and Mouth Disease in Indian Sheep and Goats. Front. Vet. Sci 2020; 7: 356.
- 24-Peng J, Yi J, Yang W, Ren J, Wen Y, Zheng H, Li D. Advances in Foot-and-Mouth Disease Virus Proteins Regulating Host Innate Immunity. Front. Microbiol 2020; 11: 2046.

- 25-Tene NST. Cameroon's adaptation to climate change and sorghum productivity. Cogent Social Sci 2022; 8: 1.
- 26-Ministère de l'élevage des peches et des industies animals. « Rapport d'activités. Décembre, Ministère de l'élevage des peches et des industies animales », 2013. MINEPIA, Yaoundé, 2015.
- 27-Lazarus DD, Schielen W, Wungak YS, Kwange D, Fasina FO. Sero-epidemiology of foot-and-mouth disease in some border states of Nigeria. African Journal of Microbiology Research 2012; 6: 1756-1761.
- 28-Thrusfield M. Veterinary epidemiology. 2nd Edition, Blackwell Science, Oxford, 2005; pp.117-198.
- 29-Salami I. « Détermination de l'âge par la dentition chez les petits ruminants en milieu traditionnel au Sénégal » Thèse doctorat en médecine vétérinaire, Ecole Inter- Etats des sciences et de Médecine Vétérinaire Dakar-Sénégal, p 1990 ; 133.
 - 30-Hervieu J, Colomer-Rocher P, Branca A, Delfa R. Définition des notes d'état corporel des caprins Réseaux Agrimed et FAO de recherches coopératives sur les productions ovines et caprines 1989 ; 5.
 - 31-Aklobessi KK. Collecte et exploitation des données existantes surla production animale au Togo. Tome I: - Considérations générales, organismes de promotion de l'élevage 1989 ;100.
 - 32-Ludi AB, Morris A, Gubbins S, Asfor A, Afzal M, Browning CF, Grazioli S, Foglia EA, Wilsden G, Burman A. Cross-Serotype Reactivity of ELISAs Used to Detect Antibodies to the Structural Proteins of Footand-Mouth Disease Virus. Viruses 2022; 14: 1495.

- 33-Relmy A, Romey A, Gorna K, Blaise-Boisseau S. Crise sanitaire dans l'Océan Indien: virus de la fièvre aphteuse aux Îles Maurice et Rodrigues en 2016 », Épi. San. Ani 2017; 71: 117–127.
- 34- Campbell I. Chi-squared and Fisher-Irwin tests of two-by-two tables with small sample recommendations. Stat. Med 2017; (26): 3661-3675.
- 35-Richardson JTE. The analysis of 2 x 2 contingency tables Yet again. Stat. Med 2011; 30: 890.
- 36-Abd Hatem A, Ahmed Abdul Wahid Al Anbagi N, Al-Alo KZK, Sabah Bustani G. Detection of clinical and subclinical Foot and Mouth Disease Virus in Cattle in Al-Najaf Province. Arch. Razi. Inst 2022; 77(3): 1185-1189.
- 37-Abu ElZein EME, Newman BJ, Crowther JR. Rev. élev. Méd. Vét. Pays Trop 1987; 40(1): 7-12.
- 38-Habiela M, Raouf A, Eldin N. Short Communication: Sero-survey of Anti-Foot and Mouth Disease Virus Antibodies in Sheep and Goats in Khartoum State, Sudan. 2009; 24: 61-64.
- 39-Balinda SN, Tjørnehøj K, Muwanika VB. Prevalence estimates of antibodies towards foot-and-mouth disease virus in small ruminants in Uganda. Transbound. Emerg. Dis 2009; 56 (9-10): 362-371.

Jumbo SD, Noumedem RNG, Wade A, Dah I, God-Yang L, Sevidzem SL, Mbanwi CM, Niba AT, Awah-Ndukum J. Seroprevalence and risk factors of foot-and-mouth disease in sheep and goats in the Soudan-Sahelian and Guinean regions of Cameroon. Microbes Infect Dis 2025; 6(3): 2042-2052.