

## Original article

# Detection of pneumococcal carriage among under five healthy children with multiple co-colonizing serotypes with impact of pneumococcal conjugate vaccine

Moonmoon Shormin <sup>\*1</sup>, Samira Afroz <sup>2</sup>, SM Shamsuzzaman <sup>3</sup>, Negar Sultana <sup>4</sup>

1- Department of Microbiology, Shaheed Monsur Ali Medical College, Dhaka, Bangladesh.

2- Department of Microbiology, Shahab Uddin Medical College, Dhaka, Bangladesh.

3- Department of Microbiology, Dhaka Medical College, Dhaka, Bangladesh.

4- Department of Pathology with Microbiology, Dhaka Dental College, Dhaka, Bangladesh.

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## ABSTRACT

**Background:** The distribution of serotypes causing invasive pneumococcal diseases (IPD) are diverse, limiting the proportion of IPD cases pneumococcal conjugate vaccine (PCV) can prevent. More studies are needed to estimate the rate of pneumococcal acquisition and serotype replacement following introduction of PCV. **Methods:** The study was conducted in the Department of Microbiology of a tertiary hospital. Nasopharyngeal swabs were collected among 200 under five children from Pediatric outpatient department. *Streptococcus pneumoniae* (*S. pneumoniae*) were isolated and identified by culture, Gram staining, biochemical test and polymerase chain reaction (PCR). An initial screening of the nasopharyngeal swabs were done by the primer *cpsA* to identify the pneumococci by monoplex PCR. Then primers that are serotype specific were used for serotyping by multiplex PCR with positive and negative control. **Results:** Out of 200 nasopharyngeal (NP) swabs, 67 (33.50%) were positive by culture and 92 (46%) were positive by PCR for *S. pneumoniae*. Out of 200 children, 90 (45%) received PCV and 110 (55%) were not vaccinated. Among vaccinated children, 3 (12%) *S. pneumoniae* were detected in fully vaccinated children and 19 (29.23%) *S. pneumoniae* were detected in partially vaccinated children by culture. In case of PCR, 4 (16%) *S. pneumoniae* were detected in fully vaccinated children and 25 (38.46%) *S. pneumoniae* were detected in partially vaccinated children. Among not vaccinated children, 45 (40.90%) and 63 (57.27%) *S. pneumoniae* were detected by culture and PCR, respectively. The predominant serotypes were 34F, 35B, 6A, 6B, 14, 23F, 3, 19A, 19F, 4, 18C, 7F as carriage strains. **Conclusion:** Children represent a consistent population of pneumococcal-naïve individuals. So, we found detection rate of *S. pneumoniae* and serotypes in children as carrier was relatively more in unvaccinated children.

## Introduction

*Streptococcus pneumoniae* (*S. pneumonia*) is a major cause of pneumonia, meningitis, and other invasive diseases resulting in high mortality and morbidity among children under the age of five, particularly in lower income countries [1]. *Streptococcus pneumoniae* is a bacterium that

colonizes the nasopharynx of human and main source is person to person transmission [2,3]. *Streptococcus pneumoniae* colonization is often asymptomatic but may cause overt infections. It is spread by respiratory droplets and children are the main source of transmission to adults. The pathogenesis of invasive

pneumococcal diseases (IPD) begins with nasopharyngeal (NP) colonization that proceeds, often through local infection, to blood stream invasion [4]. Pneumococcal colonization is a dynamic process. One serotype is usually carried at a time, with the first strain often being carried the longest and reacquisition of the same serotype is relatively common [2]. Pneumococcal polysaccharide vaccines and pneumococcal conjugate vaccines (PCV) are two types of vaccines that are used in control of invasive diseases. Use of an effective conjugate vaccine during infancy reduced the rates of invasive pneumococcal infections and antibiotic resistance associated with serotypes [5]. However, infections caused by other serotypes that are not included in different PCV have increased [6].

Although almost all children become colonized with *S. pneumoniae* repeatedly during the first few years of life, a very small fraction of these acquisitions results in invasive disease. Global surveillance demonstrates that a limited number of serotypes cause 80% of IPD, and serotypes vary in their contribution to invasive disease. There are two components to this variation: (i) some serotypes are more commonly carried and thus have more temporal opportunity for invasion, and (ii) some serotypes are more likely to cause invasive disease with each carriage episode, perhaps because of their superior ability to overcome host defenses and penetrate the bloodstream [4]. *Streptococcus pneumoniae* can produce at least 90 capsular serotypes [7], but only a few of these cause most cases of invasive diseases [8]. Since the pneumococcal epidemiology of capsule types varies geographically and temporally, a nationwide serotypes surveillance system is vital to establishing appropriate vaccination strategies [9]. In this study we detect the pneumococcal carriage with multiple serotypes, which is essential for production of the new generation of conjugate vaccines.

## Materials and Methods

### Data collection

A cross sectional study was done from July 2016 to June 2017. Nasopharyngeal swabs were collected from healthy children aged one month to less than five years who attended the outpatient department of Dhaka Medical College Hospital, Dhaka in Bangladesh for routine immunization, child growth monitoring and nutritional advice. Children whose parents did not give consent and non-co-operative children were excluded from the study. Children with recent history of upper respiratory tract infection,

hospitalization and antibiotics usage were also excluded from the study.

### Ethical approval

This protocol was approved by Research Review Committee (RRC) of Microbiology Department of Dhaka Medical College, Dhaka in Bangladesh. Informed written consent was obtained from the parents or legal guardians of each child before sample collection.

### Sample collection procedure

Samples were collected from nasopharynx of each child by inserting a specially designed swab with a flexible shaft and a small calcium alginate tip [10]. The swabs were gently inserted into the NP area located one-half to two-thirds distance from the nostril to ear lobe, Then the swab was rotated gently and was allowed to remain for 20-30 seconds. The swab was then removed and was placed in Skimmilk-tryptone-glucose-glycerol (STGG) media, labeled and transported to the laboratory [11], then the swabs containing screw-cap bottles were vortexed on high speed for 10-20 seconds and placed at -20°C in upright position for culture and polymerase chain reaction (PCR) [12].

### Culture and isolation [12]

Specimens which were kept at -20°C were hawed at room temperature (25°C) and vortexed for 10-20 seconds. Then specimens were inoculated on blood agar using one loop (10µl) of sample. The plates were streaked into four quadrants and incubated at 37°C for 24 hours with CO<sub>2</sub> atmosphere inside a candle jar. Small, smooth and transparent colonies were seen on blood agar plate. Colonies of *S. pneumoniae* were low convex, tiny and they became flattened centrally showing the 'draughtsman form'. A narrow zone of α hemolysis was seen around the colonies. Gram positive diplococci were seen which were ovoid or lanceolate in shape and catalase negative [13]. The isolates with presumptive identification were confirmed by optochin sensitivity test, bile solubility test (Table 1) and by PCR (Figure 1).

### Procedure of PCR [14]

Samples preserved in STGG medium were brought out from freeze, kept at room temperature to demoiseure, then samples were vortexed to make a homogenous suspension. Swab sticks were removed and the vortexed specimens were taken into two micro centrifuge tubes, labeled properly, centrifuged at 10,000 X g for 10 minutes and the supernatant was discarded. The deposit was used as pellet for PCR. The

micro centrifuge tubes containing pellet were kept at -20 °C until DNA extraction.

#### ▪ **DNA extraction**

Two hundred micro litter of lytic buffer was mixed with the sample pellets and vortexed until mixed well. Then the tubes were incubated at 60°C for 3 hours. After incubation, tubes were kept in heat block at 100°C for 10 minutes for boiling. Then the tubes were immediately placed on ice for 5 minutes. After that the tubes were centrifuged at 4°C at 14000 X g for 10 minutes. Finally supernatants were taken using micropipette and used as template DNA for PCR. This DNA was kept at -20°C for future use [10].

#### ▪ **Serotyping by PCR**

The primer *cpsA* was used for targeted highly conserved gene that exists in all capsular loci thus far characterized [15]. Thirty eight specific primers were used for targeted genes specific for serotypes 1, 2, 3, 4, 5, 6A, 6B, 7F, 9V, 11A, 12F, 14, 18C, 19F, 19A, 23F, 33F, 34F and 35B. Of these serotypes 1, 2, 3, 4, 5, 6A, 6B, 7F, 9V, 11A, 12F, 14, 18C, 19F, 19A, 23F, 33F were vaccine type (VT) and serotypes 34F, 35B were non vaccine (NVT) type. Vaccine Type serotypes means a serotype included in PCV7 (serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F), PCV10 (serotypes 1, 5, and 7F added to PCV7), PCV13 (serotypes 3, 6A, and 19A added to PCV10), and PPSV23 (serotypes 2, 8, 9N, 10A, 11A, 12F, 15B, 17F, 20, 22F, and 33F added to PCV13, except for 6A). Non-vaccine serotypes are the serotypes which are not covered by PCV7, PCV10, PCV13, and PPSV23. An initial screening of the NP swabs were done by the primer *cpsA* to identify the pneumococci by monoplex PCR (Table 2a). Then primers that are serotype specific were used for serotyping by multiplex PCR with positive and negative controls [10] (Table 2b).

#### ▪ **Mixing of master mix and primer with DNA template**

Primers were mixed with Tris-EDTA (TE) buffer according to manufacturer's instruction. For each sample, total 25µl of mixture was prepared by mixing of 12.5 µl of master mix (mixture of dNTP, taq polymerase MgCl<sub>2</sub> and PCR buffer), 2µl of forward primer, 2µl of revers primer (Promega Corporation, USA), 2µl of DNA template and 6.5 µl of sterile distilled water in a micro centrifuge tube. For multiplex PCR, total 25 µl of mixture was prepared by mixing of 12.5 µl of master mix, 1µl of each forward primer, 1µl of revers primer, 2µl of DNA template and nuclease free water in a micro centrifuge tube. After a brief vortex, the tubes were centrifuged in a micro centrifuge for few seconds [10].

Amplification in thermal cycler (Eppendorf AG, Master cycler gradient, Hamburg, Germany):

Melting temperature of the primer depends on base pair and calculated using the following formula:

- Up to 20 bp, the melting temperature is (°C) = 4 (G+C) +2 (A+T)-5.
- More than 30 bp, the melting temperature is (°C) = 4(G+C) +2 (A+T)+ 35-2n.  
(n= total number of base pair)

Polymerase chain reaction consisted of preheat at 94°C 10 minutes followed by 36 cycles of denaturation at 94 °C for 30 seconds, annealing at 52 °C for 40 seconds, extension at 72 for one minute with final extension at 72 for 10 minutes. Annealing temperature varies with GC contents of primers.

#### ▪ **Polymerase chain reaction primer selection**

The selection of PCR primers was based on the published works [15,16] (Table 2).

#### ▪ **Agarose gel electrophoresis**

Polymerase chain reaction products were detected by electrophoresis on 2% agarose gel. Gel was prepared with 1X TBE buffer (Tris Borate EDTA) For 2% agarose gel preparation, 0.25 gram of agarose powder (LE, Analytical grade, Promega, Madison, USA) was mixed with 12.5 ml TBE buffer. After mixing thoroughly it was boiled using microwave oven for few minutes to dissolve the agarose completely. Gel was cooled to 60-70 °C and a comb was placed in gel tray and the agarose gel was poured into the tray and was allowed to stand for 30 minutes for solidification. After solidification of gel, comb was removed from the tray. Six µl of amplicon was mixed with 1µl loading dye on a piece of para film and then the mixture was loaded into the well of agarose gel. Two µl DNA ladder was mixed with 1 µl of loading dye and was loaded into one well of agarose gel. Gel containing amplicons and DNA ladder were then placed on the electrophoresis tank having 1X TBE buffer for 35 minutes at 100 volts. Positive control and negative control were also loaded in separate well.

#### ▪ **Staining and de-staining of the gel**

After electrophoresis, the gel was stained with staining buffer for 30 minutes. It was then de-stained with distilled water for 15 minutes.

#### **Data analysis**

The results of the study were recorded systematically. Data analysis was done by using 'Microsoft Office Excel 2007' program and according to the objectives of the study. Results were presented in the form of tables and figures. The test of

significance was calculated by using Z test and  $X^2$  test.  $P$  value  $<0.05$  was taken as minimal level of

significance and  $p$  value  $<0.001$  was taken as highly significant.

**Table 1.** Identification of *S.pneumoniae* and its differences with Viridans streptococci.

Characteristic	<i>S.pneumoniae</i>	Viridans streptococcus
Colonies	Flattened, draughtsman	Convex
Effect on blood agar	Narrow zone of $\alpha$ haemolysis	Narrow zone of $\alpha$ haemolysis
Optochin sensitivity	+ve	-ve
Bile solubility	+ve	-ve

**Table 2a.** List of selected *cpsA* primer with annealing temperature and product size.

Name of gene	Oligonucleotide sequence (5'-3')	Annealing temperature <sup>o</sup> C	Product size
<b>cpsA-f</b>	GGT GTT CTC TAT CCT TGT CAG CTC TGT GTC GCT C	56	657
<b>cpsA-r</b>	GTG TGA ATG GTC GAA TCA ACT CTA TAA ATG CC		
<b>1wzy-f</b>	GGA GAC TAC TAA ATT GTA ATA CTA ACA CAG CG	48	280
<b>1wzy-r</b>	CAA GGA TGA ATA AAG TAA ACA TAT AAT CTC		
<b>3capB-f</b>	TTG TTT TTT GTC TTT ATT CTT ATT CGT TGG	52	818
<b>3capB-r</b>	TAC TGA GAA CCT TCT GCC CAC CTT AGT TGC		
<b>4wzy-f</b>	CTG TTA CTT GTT CTG GAC TCT CGT TAA TTG G	56	430
<b>4wzy-r</b>	GCC CAC TCC TGT TAA AAT CCT ACC CGC ATT G		
<b>6Bwzy-f</b>	CGA CGT AAC AAA GAA CTA GGT GCT GAA AC	52	220
<b>6Bwzy-r</b>	AAG TAT ATA ACC ACG CTG TAA AAC TCT GAC		
<b>14cpsH-f</b>	GTC TGT TTATTC TAT ATA CAA AGA GGC TCC	51	268
<b>14cpsH-r</b>	GCA TTG CTA CAA TCG CTA TAC TAG ATA TGC		
<b>18CwciY-f</b>	GCA TCT GTA CAG TGT GCT AAT TGG ATT GAA G	52	354
<b>18Cgct-r</b>	CTT TAA CAT CTG ACT TTT TCT GTT CCC AAC		
<b>19FcpsI-f</b>	CAC CTA ATT TTA ATA CTG AGG TTA AGA TTG C	48	408
<b>19FcpsI-r</b>	CAT AGG CTA TCA GAA TTT TAA TAA TAT CTT GC		
<b>19AcpsK-f</b>	GTT AGT CCT GTT TTA GAT TTA TTT GGT GAT GT	51	478
<b>19AcpsK-r</b>	GAG CAG TCA ATA AGA TGA GAC GAT AGT TAG		
<b>23FcpsG-f</b>	GTA ACA GTT GCT GTA GAG GGA ATT GGC TTT TC	55	384
<b>23FcpsG-r</b>	CAC AAC ACC TAA CAC ACG ATG GCT ATA TGA TTC		

**Table 2b.** List of primers selected for serotyping with annealing temperature and product size.

Primer Pair	Oligonucleotide sequence (5'-3')	Annealing temperature°C	Product Size
2-f	GTC ATT GTT ACG ATT AGT TTC GAT AGT TGA GG	51	381
2-r	AAT TCA ATT CCT AAG TCC TCT TCC ATA AAC TC		
6A-f	AAT TTG TAT TTT ATT CAT GCC TAT ATC TGG	54	250
6A-r	TTA GCG GAG ATA ATT TAA AAT GAT GAC TA		
7F-f	CTA TCT CAG TCA TCT ATT GTT AAA GTT TAC GAC GGG A	54	260
7F-r	GAA CAT AGA TGT TGA GAC ATC TTT TGT AAT TTC		
9V-f	CTT CGT TAG TTA AAA TTC TAA ATT TTT CTA AG	54	753
9V-r	GTC CCA ATA CCA GTC CTT GCA ACA CAA G		
11A-f	GGA CAT GTT CAG GTG ATT TCC CAA TAT AGT G	54	463
11A-r	GAT TAT GAG TGT AAT TTA TTC TTC CAA CTT CTC CC		
12F-f	GCA ACA AAC GGC GTG AAA GTA GTT G	54	376
12F-r	CAA GAT GAA TAT CAC TAC CAA TAA CAA AAC		
33F-f	GAA GGC AAT CAA TGT GAT GAT TGT GTC GCG	54	338
33F-r	CTT CAA AAT GAA GAT TAT AGT ACC CTT CTT CTA C		
34-f	GCT TTT GTA AGA GGA GAT TAT TTT CAC CCA AC	54	408
34-r	CAA TCC GAC TAA GTC TTC AGT AAA AAA CTT TAC		
35-B-f	GAT AAG TCT GTT GTG GAG ACT TAA AAA GAA TG	54	677
35-B-r	CTT TCC AGA TAA TTA CAG GTA TTC CTG AAG CAA G		

## Results

Nasopharyngeal swabs from total 200 under five children were tested (**Table 3**) [in this study relations between pneumococci carriage, serotyping, vaccination and age were not seen], among them, 67(33.50%) were positive for *S. pneumoniae* by culture (**Table 4**) and 92 (46%) were positive by PCR using *cpsA* gene specific primer (**Table 5, Figure 1**). Among 92 PCR positive NP swabs, 19 different serotypes were seen. Among 92 PCR positive NP swabs, 70 (76.09%) were serotypes positive and 22 (23.91%) were serotype negative (**Figure 2**). Among them more detected serotypes were 34F (17.39%), 35B (13.04%), 6A (11.96%), 6B (11.96%), 14 (7.61%) and 23F 4 (5.43%). Less detected serotypes were 3 (3.26%), 19A (2.17%), 19F (2.17%), 4 (1.09%), 18C (1.09%) and 7F (1.09%) (**Figure 3**). Out of 200 children, 90 (45%) were received (PCV) and 110 (55%) were not vaccinated. Among vaccinated children, 3 (12%) *S. pneumoniae* were detected in fully vaccinated children (3 doses as recommended in

Bangladesh; mentioned in the related table) and 19 (29.23%) *S. pneumoniae* were detected in partially vaccinated children by culture. In case of PCR, 4 (16%) *S. pneumoniae* were detected in fully vaccinated children and 25 (38.46%) *S. pneumoniae* were detected in partially vaccinated children. Among non-vaccinated children, 45 (40.90%) and 63 (57.27%) *S. pneumoniae* were detected by culture and PCR respectively (**Table 6**). The difference between fully vaccinated and non-vaccinated proportion is statistically significant ( $p < 0.001$ ) and the difference between fully vaccinated and partially vaccinated proportion is statistically significant ( $p < 0.001$ ). Among 4 fully vaccinated PCR positive cases, serotypes (mentioned in methodology) were detected among 3 fully-vaccinated children, in the rest one vaccinated child serotypes was not detected as all the serotype specific primers were not used in this study and detected serotypes were 34F (1) and 35B (1) and 6B (1); in the rest one vaccinated child serotypes was not detected as all the serotype specific primers were not used in this study. Among 25 PCR positive

partially-vaccinated children, serotypes were detected in 18 children and detected serotypes were 34F (7), 35B (3), 6B (3), 14(3), 23(2) [here mentioned only numerical number which were found in individual type of vaccinated children, percentage(%) of total detected serotypes were mentioned in result paragraph] (Figure 4). Among 63 PCR positive non-vaccinated children, serotypes were detected in 51 children and the detected serotypes were 3 (3), 4 (1), 6A (11), 6B (7), 7F (1), 14 (4), 18C (1), 19A (2), 19F (2), 23F (3), 34F (8), 35B (8) (Table 7), (Figure 5); [here mentioned only numerical number which were found in individual type of vaccinated children, percentage(%)of total detected serotypes were mentioned in result pharagraph .

**Table 3.** Results of culture and PCR in different age groups (n=200).

Age groups	Culture positive n (%)	PCR positive n (%)
1month-12 months (n= 40)	13(32.50)	21(52.50)
13months-36 months (n= 65)	29(44.62)	38(58.46)
37months- ≤ 60 months (n= 95)	25(26.32)	33(34.74)
<b>Total</b>	<b>67</b>	<b>92</b>

N= Total number of samples, n= Total number of positive cases % were calculated according to the row.

**Table 4.** Results of culture for *S. pneumoniae* from NP swabs (n=200).

Methods	Nasopharyngeal swabs n (%)
Culture positive	67(33.50)
Culture negative	133 (66.5%)
<b>Total</b>	<b>200 (100%)</b>

**Table 5.** Results of PCR for *S. pneumoniae* from NP swabs (n=200).

Methods	Nasopharyngeal swabs n (%)
PCR positive	92(46%)
PCR negative	108 (54%)
<b>Total</b>	<b>200 (100%)</b>

**Table 6.** Distribution of *S. pneumoniae* among study population in relation to vaccination status (n=200).

Vaccination status	Culture positive n (%)	PCR positive n (%)
Fully vaccinated* (n=25)	3 (12)	4 (16)
Partially vaccinated** (n=65)	19(29.23)	25(38.46)
Not vaccinated (n=110)	45(40.90)	63(57.27)
<b>Total</b>	<b>67</b>	<b>92</b>

\*Vaccinated with 3 doses of PCV

\*\*Vaccinated with 1 or 2 doses of PCV.

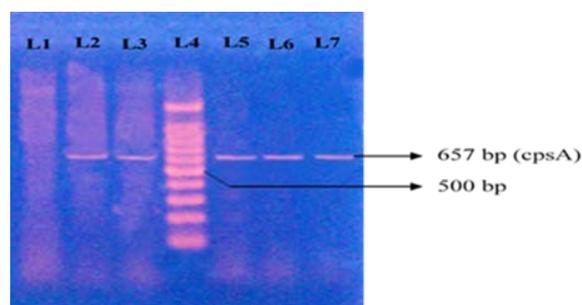
**Table 7.** Distribution of detected serotypes among vaccinated, partially vaccinated and non-vaccinated children in PCR positive children (n=92).

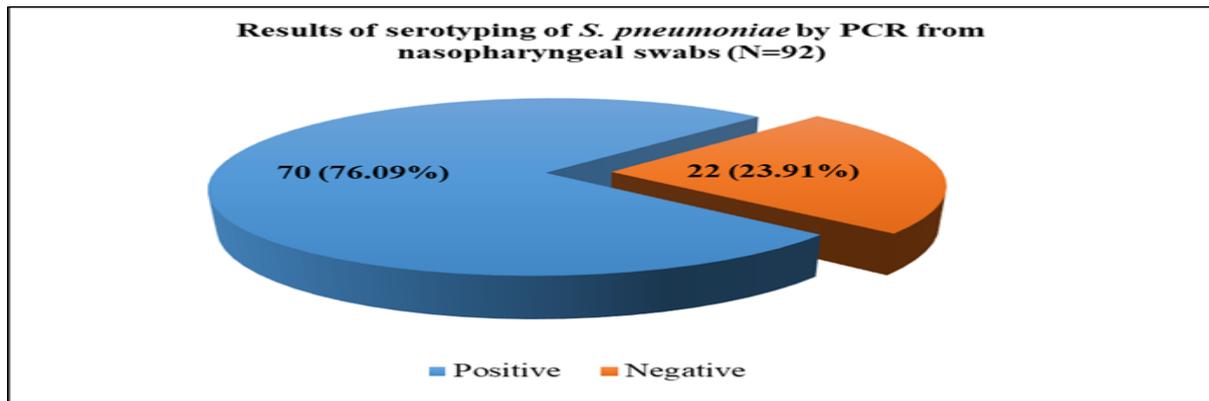
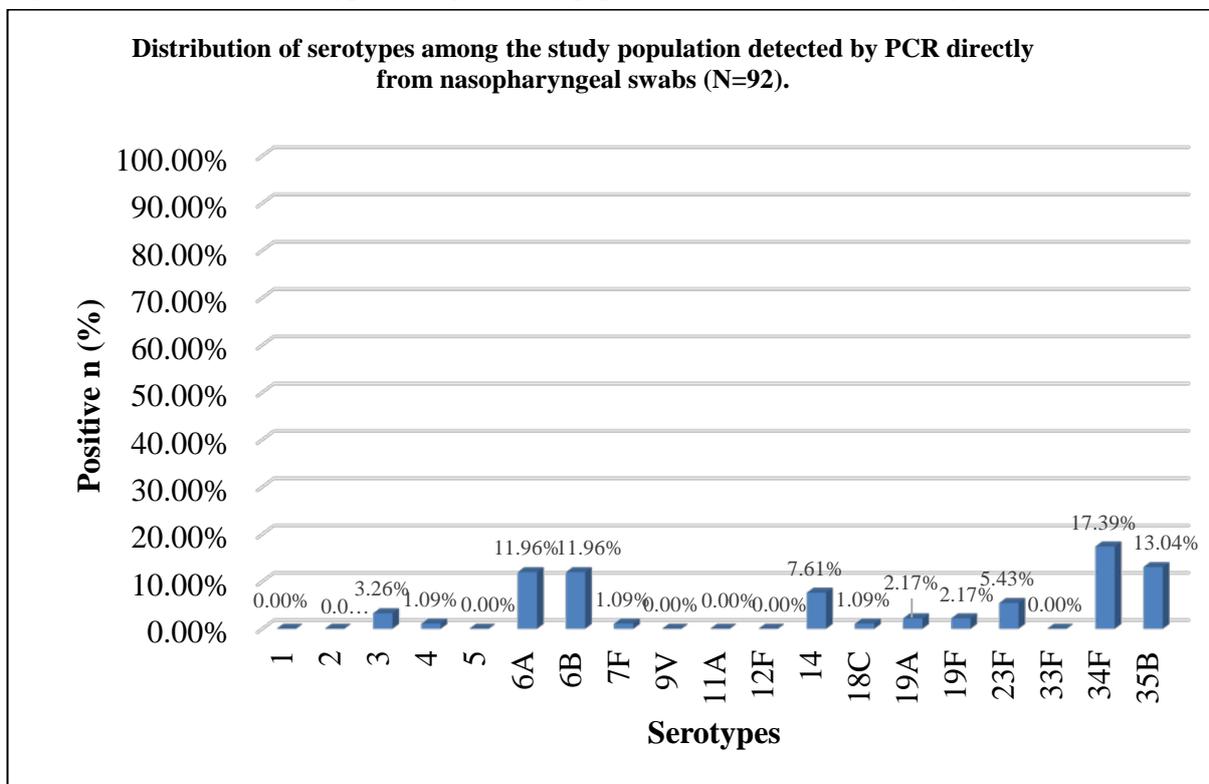
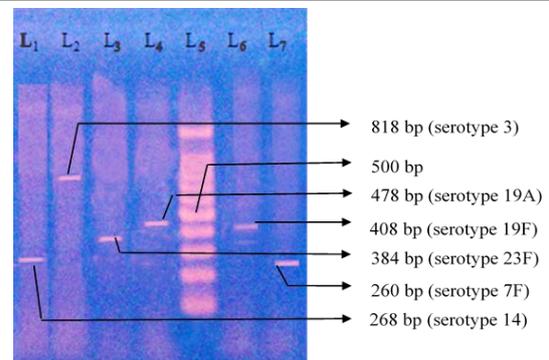
Vaccination status	Detected serotypes(n)
Fully vaccinated (n=4)	*34F(1), *35B (1), 6B (1)
Partially vaccinated (n=25)	*34F(7), * 35B(3),6B (3), 14 (3), 23F(2)
Not-vaccinated (n=63)	3(3), 4(1), 6A(11), 6B(7), 7F(1), 14(4), 19A(2), 19F(2), 23F(3), *34F(8), *35B(8)

VT=3, 4, 6A, 6B, 7F, 14, 18C, 19A, 19F, 23F, \*NVT=34F, 35B

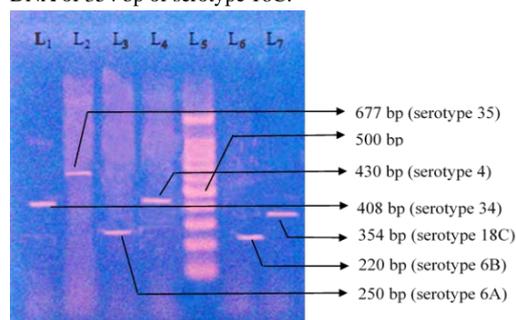
VT-Vaccine Type serotypes: a serotype included in PCV7 (serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F), PCV10 (serotypes 1, 5, and 7F added to PCV7), PCV13 (serotypes 3, 6A, and 19A added to PCV10), and PPSV23 (serotypes 2, 8, 9N, 10A, 11A, 12F, 15B, 17F, 20, 22F, and 33F added to PCV13, except for 6A). NVT-Nonvaccine Type serotypes: serotypes which are not covered by PCV7, PCV10, PCV13, and PPSV23.

**Figure 1.** Photograph of amplified *cpsA* gene of *Streptococcus pneumoniae*. Lane 1 shows negative control with DNA of *Staphylococcus aureus* ATCC25923, Lane 2, 3, 5, 6, 7 shows amplified DNA of 657 bp of *cpsA* gene. Lane 4 shows hundred bp DNA ladder.



**Figure 2.** Proportion of detected serotype positive and negative NP by PCR.**Figure 3.** Distribution of serotypes among the study population detected by PCR directly from NP swabs (n=92).**Figure 4.** Photograph of gel electrophoresis of detected serotypes. Lane 1: amplified DNA of 268 bp of serotype 14, Lane 2: amplified DNA of 818 bp of serotype 3, Lane 3: amplified DNA of 384 bp of serotype 23F, Lane 4: amplified DNA of 478 bp of serotype 19A, Lane 5: hundred bp DNA ladder, Lane 6: amplified DNA of 408 bp of serotype 19F and Lane 7: amplified DNA of 260 bp of serotype 7F.**Figure 5.** Photograph of gel electrophoresis of detected serotypes. Lane 1: amplified DNA of 408 bp of serotype 34, Lane 2: amplified DNA of 677 bp of serotype 35, Lane 3: amplified DNA of 250 bp of serotype 6A, Lane 4: amplified DNA of 430 bp of serotype 4, Lane 5: hundred bp DNA ladder, Lane 6:

amplified DNA of 220 bp of serotype 6B, and Lane 7 : amplified DNA of 354 bp of serotype 18C.



## Discussion

*Streptococcus pneumoniae* is a bacterium that colonizes the nasopharynx of human, it's most common cause of pneumonia and other IPD. The capsular polysaccharide, a well-known virulence factor and the serotype determinant of pneumococci that prevents opsonization and phagocytosis of *S. pneumoniae* [17]. Pneumococcal capsular vaccines protect against disease and nasopharyngeal carriage due to the serotypes included in the vaccine formulations [18]. Nevertheless, surveillance of serotype prevalence patterns is very important since the serotypes responsible for invasive disease can change over time [19,20]. The surface capsular polysaccharide of *S. pneumoniae* provokes a type-specific protective immune response and serves as the basis for serotyping of these organisms; currently more than 90 different pneumococcal serotypes have been identified. Serotypes 6, 14, 18, 19, and 23 are the most prevalent, accounting for between 60 and 80 percent of infections depending upon the area of the world.

Ninety capsular serotypes of *S. pneumoniae* exist, and the prevalence of serotypes differs according to age, region, and time of the surveillance. The 90 serotypes differ in virulence; a minority of serotypes is involved in most of invasive pneumococcal diseases and antimicrobial resistances. Serotypes are classified into VT and (NVT. Vaccine serotype means a serotype included in PCV7 (serotypes 4, 6B, 9V, 14, 18C, 19F, and 23F), PCV10 (serotypes 1, 5, and 7F added to PCV7), PCV13 (serotypes 3, 6A, and 19A added to PCV10), and PPSV23 (serotypes 2, 8, 9N, 10A, 11A, 12F, 15B, 17F, 20, 22F, and 33F added to PCV13, except for 6A). Nonvaccine serotype is the serotype which is not covered by PCV7, PCV10, PCV13, and PPSV23 [9].

In the present study, a total of 200 NP swabs were processed for culture and PCR. Out of 200 NP swabs, 67 (33.50%) were culture positive and 92

(46%) were PCR for *S. pneumoniae*. In the present study, PCR base detection of carriage was higher than culture (46% vs 33.50%). Some previous studies [21,22] reported that PCR significantly increased the detection of carriage of *S. pneumoniae* compared to culture. The results of the previous studies and the results of the present study showed that PCR based detection is higher in comparison to culture. *Streptococcus pneumoniae* isolation from NP swab by culture was less sensitive which might be due to the fastidious nature of the organism and low load of the organism with co-colonization that can result in false negative results [23]. Polymerase chain reaction can detect bacterial DNA even if the numbers of organism is too low to grow in culture [24] and can detect from non-viable organisms after treatment with antibiotics [25].

In the present study, among 92 PCR positive NP swabs, 19 different serotypes were seen by PCR. Out of 19, 17 (1, 2, 3, 4, 5, 6A, 6B, 7F, 9V, 11A, 12F, 14, 18C, 19A, 19F, 23F, 33F) were vaccine types (VT) and 2 (34F, 35B) were non vaccine types (NVT). Among 70 serotype positive NP swabs, 72 different serotypes were detected. More detected serotypes were 34F (17.39%), 35B (13.04%), 6A and 6B (11.96%), 14 (7.61%), 23F (5.43%) and less detected serotypes were 3 (3.26%), 19A and 19B (2.17%), 4, 7F and 18C and were all was 1.09%. Serotypes 1, 2, 5, 9V, 11A, 33F were not found in the present study. In the present study the detected serotypes were similar to other studies conducted in Bangladesh in different period of times. Two studies<sup>26, 27</sup> detected serotypes (14, 7F, 6A, 6B, 18C, 19A, 23F) from invasive cases and from both meningitis (6, 7, 14, and 18) and carriage cases (6, 19, 23, 34, 35, and 14).

In the present study serotype 1, 2, 5, 9V, 11A, 33F were not found. The reasons might be, pneumococcal colonization is a dynamic process. One serotype is usually carried at a time, with the first strain often being carried the longest and reacquisition of the same serotype is relatively common [2], besides, serotypes or serogroups carried for a long period would be recovered more frequently from nasal swabs than those carried only transiently, and invasive diseases is more likely to occur soon after the acquisition and thus, is less well associated with the period of colonization [26]. In the present study, carriage rate was significantly higher in non-vaccinated children than fully-vaccinated children ( $p < 0.001$ ) and also significantly higher in partially-vaccinated children than fully-vaccinated children ( $p < 0.001$ ).

The findings of the present study might be due to the fact that introduction of PCV reduced the carriage rate in vaccinated children and reduced carriage of VT pneumococci accompanied by an increased NVT [28]. Some previous studies analyzed the impact of pneumococcal vaccination on *S. pneumoniae* carriage rate and serotype distribution [29-31]. They reported that overall carriage rate in residents decreased following vaccination, carriage rates of all age group were lower at first and second post vaccination surveys than at the pre vaccination survey. Some previous studies showed that the prevalence of vaccine type (VT) carriage decreased after PCV vaccination [29,31]. **Isaacman et al.** and **Flasche et al.** reported that reduction in carriage of VT pneumococci due to PCV vaccination is often accompanied by an increase in the carriage of NVT pneumococci and to a lesser extent, an increase in the incidence of IPD caused by NVT pneumococci [32,33].

#### Limitations

Due to time and resource constraint, all the VT and NVT serotypes could not be detected.

#### Conclusion

Detection rate of NP carriage of *S. pneumoniae* was relatively more and the predominant serotypes were 34F, 35B, 6A, 6B, 14, 23F, 3, 19A, 19F, 4, 18C, 7F as carriage strains. These findings highlight the importance of monitoring pneumococcal nasopharyngeal carriage and serotypes distribution for the future pneumococcal conjugate vaccine production.

#### Recommendation

We recommend that the continuous survey for detecting pneumococcal carriage with different serotypes in children can be useful and improving the ability to diagnose invasive Pneumococcal disease burden and implications for vaccine policy.

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#### Informed consent

Informed written consent was obtained from the parents or legal guardians of each child before sample collection.

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